

Research Article

RP-HPLC Method Development and Validation for the Determination of Ezetimibe Using Design of Experiments Approach

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Abstract

The present research aims to develop and validate a simple and accurate RP-HPLC method for the determination of Ezetimibe by using the Design of Experiments Approach. This approach was useful for multivariate optimization of the method. The critical method parameters (CMPs) were optimized using the Box-Behnken design. Minitab software was equipped for the study. Chromatographic separation was done on Phenomenex C18 column with specifications 150 mm × 4.6 mm × 5 μm at 30°C. The predicted and optimized data from the software consisted of mobile phase 0.02 N Ortho phosphoric acid (OPA) and Acetonitrile (53:47% v/v), pumped at a flow rate of 0.96 ml/min brought the desirability function of 1. The UV detector was adjusted at 232.6 nm. The developed method shows linearity with a correlation coefficient of 0.999. The optimized chromatographic method was validated as per the guidelines of ICH Q2 (R1). The stability of drug was forcibly studied under different stress conditions.

Keywords: Ezetimibe; Design of Experiments Approach; Box-Behnken design; ICH Q2 (R1).

Introduction

Ezetimibe [1,2] is marketed under the brand name Zetiheal, which is approved for the management of hypercholesterolemia. Generally, this drug decreases the absorption of cholesterol and phytosterol via small intestine without disturbing the absorption of fat-soluble vitamins and minerals by that means it lowers blood cholesterol levels. The IUPAC name of the compound is (3R,4S)-1-(4-fluorophenyl)-3-[(3S)-3-(4-fluorophenyl)-3-hydroxypropyl]-4-(4-hydroxyphenyl)azetidin-2-one and the chemical structure of the compound is shown in Figure 1. After detailed literature review of Ezetimibe, a few methods are reported based on a variety of techniques such as UV-spectroscopy,

Liquid chromatographic-tandem mass spectrometric (LC-MS/MS) methods and HPLC Methods [3-5]. None of the reported analytical methods describes a simple HPLC method for studying the effect of stress on pharmaceutical dosage forms of Ezetimibe. Hence the present work was focused on the development and validation of the estimation of Ezetimibe by Analytical Quality by Design (AQbD) approach with the help of Minitab software. This approach helps in understanding the empirical relationship between one or more measured responses and several independent variables in the form of a polynomial equation. Mapping of those responses related to the experimental domain helps in developing an optimized method. Optimization of the method for the present research was performed with the help of the Box-Behnken design.

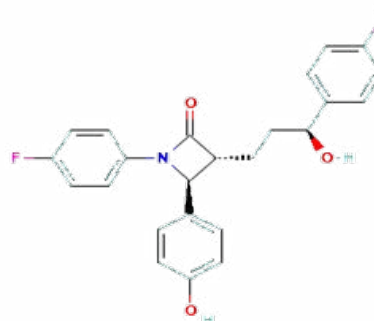


Figure 1: Chemical structure of Ezetimibe

Materials and Methods

Ezetimibe is obtained from Spectrum Labs. Phosphate buffer, Acetonitrile, Potassium dihydrogen orthophosphate buffer, Methanol, and ortho phosphoric acid are brought from Rankem. The formulation Zetiheal (Ezetimibe 10 mg) was purchased from the local market. Water HPLC 2695 system with photodiode array detector integrated with Empower 2 software is used for HPLC study. Minitab 21.2.0.0 was aided for the production of 2D contour plots and 3D surface plots.

Preparation of stock, standard and test solutions

Stock solution 100 µg/ml of Ezetimibe reference standard was prepared by transferring 10 mg, accurately weighed drug, into a 100 ml volumetric flask and adding 20 ml 0.02 N ortho phosphoric acid:acetonitrile (53:47% v/v). The mixture was sonicated for about 2 minutes to dissolve the Ezetimibe and then the solution was diluted to volume with the same solvent mixture. A standard solution of 10 µg/ml was prepared by diluting 1 ml standard stock solution to 10 ml, in a volumetric flask, with the same solvent mixture. To make stock solution 100 µg/ml for assay, 20 tablets were weighed and mixed. An aliquot of powder equivalent to the weight of 10 mg was accurately weighed and transferred to 100 ml volumetric flask. Add 20 ml of 0.02 N ortho phosphoric acid:acetonitrile (53:47% v/v) to the flask and the mixture was sonicated for 10 minutes in a sonicator. The contents of the flask were then diluted to the volume with the same solvent mixture. This solution (10 ml) was filtered through a nylon syringe filter of 0.45 µm. To make test solution (10 µg/ml) for assay, take 1 ml test stock solution into 10 ml volumetric flask and diluted to volume with 0.02 N ortho phosphoric acid:acetonitrile (53:47% v/v).

Initial chromatographic conditions screening

For initial chromatographic conditions selection like organic modifier, organic phase, and flow rate, a 23 Factorial design consisting of 3 factors at 2 levels was selected [6]. The selected 23 factorial design results in 8 trial runs suggest various combinations for the factors chosen.

Optimization of the method by response surface methodology

Many response surface design models are used for the optimization of methods like Central composite, Doehlert, and

Box–Behnken. Among all the Box–Behnken Design is superior because it requires lesser test runs, is rotatable, and does not contain any outliers of the cubic region. In the current investigation, the Box–Behnken design was used for the method optimization of Ezetimibe by RP-HPLC because the design provides three levels for each factor and requires lesser runs in the three-factor case than the Central composite and Doehlert design [7].

Method validation

The validation of the final optimized analytical method was performed as per ICH Q2 (R1) guidelines for specificity, system suitability, linearity, accuracy, precision, the limit of detection, the limit of Quantitation and robustness.

Forced degradation studies

The drug was subjected to various stress conditions as mentioned in ICH Q1A (R2) guidelines to understand whether the developed method was stability indicating.

Results and Discussion

Analytical QbD-assisted method development

Initial chromatographic conditions screening: A 23 Factorial design comprising three factors at two levels was selected for the experimental plan initially to select the appropriate percent of organic content in the mobile phase, organic modifier, and flow rate which majorly affect the selectivity. The percent of organic content of the mobile phase was chosen between 30-50, Methanol and Acetonitrile were chosen as organic solvents since they were most commonly used in RP-HPLC. The flow rate is adjusted between 0.8 to 1. Likewise, the factors and the levels selected for the screening design were shown in Table 1. The selected 23 factorial designs resulted in [8] trial runs signifying various combinations for the factors chosen were presented in Table 2. The responses selected were retention time, theoretical plates, and asymmetry [9,10].

Table 1: Factors and levels chosen for 23 factorial design of Ezetimibe

Factors	Levels
% Organic content of the mobile phase	30/50
Organic Modifiers	Methanol/Acetonitrile
Flow Rate	0.8/1

Table 2: Trial runs of 23 factorial design with responses

Run Order	% Organic Composition	Flow Rate	Organic Modifier	Retention Time	Theoretical Plates	Asymmetry
1	30	0.8	ACN	2.289	8647	1.2
2	30	0.8	Methanol	2.457	7988	1.2
3	50	1	ACN	2.247	8784	1.1
4	50	0.8	Methanol	2.468	7968	1.0
5	30	1	Methanol	2.472	8123	1.2
6	30	1	ACN	2.291	8562	1.2
7	50	0.8	ACN	2.325	8216	1.1
8	50	1	Methanol	2.321	8869	1.2

Statistical analysis of 2³ factorial design experimental data by Minitab software: Analysis of variance (ANOVA) was applied to study the significance of the model shown in Table 3. From the table, it is seen that the Model F-values of 4521.60, 549.69, and 15.45 for retention time, theoretical plate, and asymmetry respectively imply the model is significant. Values of $P < 0.05$ A, B, and C are significant model terms for asymmetry with $P < 0.05$. The significance of the terms A, B, and C indicates that the initial

chromatographic conditions selected have a greater influence on the responses. The responses obtained were feedback to the Minitab software and the cube plots for retention time, theoretical plates, and asymmetry was drawn as shown in Figure 2. From the 23 factorial designs, based on the cube plots for the responses the initial chromatographic conditions selected for further study were acetonitrile, 47% organic content, and 0.96 flow rate at which retention time is less, theoretical plates are more and asymmetry is less.

Table 3: ANOVA for the responses by 23 factorial model

ANOVA for selected Factorial model							
Analysis of variance (Type-III of the Partial sum of squares)							
Response	Source	Sum of squares	df	Mean square	F value	p value	Inference
Retention Time	Model	23.71	4	5.71	4521.60	<0.0001	Significant
	A-OP	0.023	1	0.023	18.21	0.021	
	B-FR	1.81	1	1.81	1403.19	<0.0001	
	C-OM	20.15	1	20.15	15380.46	<0.0001	
	AB	1.70	1	1.70	1301.21	<0.0001	
	Residual	3.932E-001	3	1.311E-001			
Theoretical Plates	Model	8.221E+005	4	2.055E+007	549.69	<0.0001	Significant
	A-OP	5.956E+003	1	5.957E+0 06	159.61	0.001	
	B-FR	5.781E+003	1	5.781E+006	154.93	0.001	
	C-OM	6.715E+005	1	6.715E+007	1799.55	<0.0001	
	AB	3.321E+005	1	3.321E+006	88.91	0.002	
	Residual	1.121E+003	3	37322.1			
Asymmetry	Model	0.35	3	0.11	15.45	0.010	Significant
	A-OP	0.081	1	0.080	11.12	0.027	
	B-FR	0.083	1	0.085	11.71	0.023	
	C-OM	0.21	1	0.21	26.51	0.005	
	Residual	0.027	4	7.362E-001			

df - Degree of Freedom, OP – Organic Phase, FR – Flow Rate, and OM – Organic Modifier

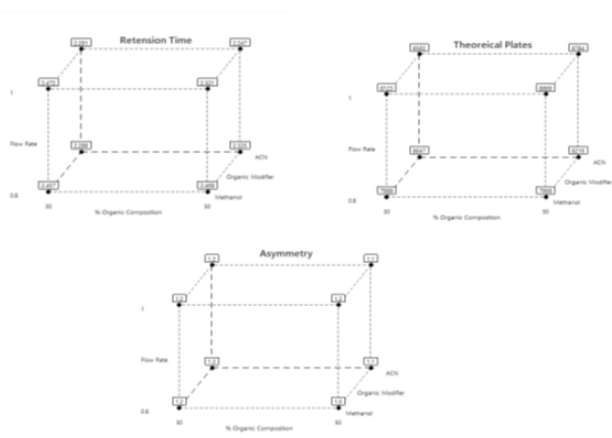


Figure 2: Cube plots of Retention Time, Theoretical Plate and Asymmetry for 23 Factorial design

Optimization by Box-Behnken design

AQbD method encompasses identifying Critical method parameters (CMP) and Critical quality attributes (CQA)

with risk assessment and producing design space. In the present study Critical Method Parameters selected were the percentage of organic content in the mobile phase, organic modifier and flow rate. The CQA's selected were retention time, theoretical plates and asymmetry. So, Box-Behnken Design was used to optimize these parameters which were varied over three level (high, mid and low)10. Different ranges of three parameters 30%-50% organic phase, organic modifiers–Acetonitrile/methanol and flow rate of 0.8-1.0 ml/min were considered. A 3-factor 3-level Box Behnken Design was developed [11]. This study design of 17 experimental runs was generated and analyzed by Minitab software as shown in Table 4.

Statistical analysis of experimental data by Minitab software

Application of the ANOVA was done to study the significance of the model for the 3 responses which is given in Table 5. The retention time, the Model F-value of 67.13 indicates the model is significant. There are only 0.01%

conditions that an F-value this large could happen due to noise. P values less than 0.0500 show model terms are significant. In this case, A, B, and C are the significant model terms. 2D Contour plots were studied to visualize the effect of factors and their interactions on the responses using the Minitab software. In the 2D Contour plots of retention time shown in Figure 3, it was found that at middle organic phase content, higher flow rate and acetonitrile as an organic modifier have less retention time. From the ANOVA data for theoretical plates shown in Table 6, the Model F-value of 2.59 suggests the model is not significant and there is a 9.5% chance that an F-value was large happen due to noise. In this case, only A is a significant model term. In the screening studies, at acetonitrile theoretical plates are more when compared to methanol and the model was found to be significant. Hence acetonitrile was selected for optimization study. To study the effect of significant term on Theoretical Plates, a 2D contour plot was analyzed using Minitab software from the 2D Contour plot of theoretical plates shown in Figure 4, it was found that at a higher organic phase content, flow rate and acetonitrile of theoretical plates is more. From the ANOVA for the asym-

metry shown in Table 7, the Model F-value of 5.31 implies the model is significant. There is only a 1.89% chance that an F-value this large could occur due to noise. P values less than 0.0500 indicate model terms are significant. In this case, AB and C2 are significant model terms. To study the effect of significant terms AB and C2 on asymmetry, a 2D contour plot was analyzed using Minitab software. From the above 2D Contour plot of asymmetry shown in Figure 5, it was found that at a higher organic phase content and acetonitrile the value of asymmetry is less. From the fit statistical parameters found from ANOVA, it was obtained that the predicted R^2 value of retention time 0.8909 is in realistic agreement with the adjusted R^2 0.9274 i.e., the difference is less than 0.2. A negative predicted R^2 value of tailing factor -0.9613 suggests that the overall mean may be a better predictor of the response than the current model. Adequate Precision finds the signal-to-noise ratio. A ratio greater than 4 is desirable and the obtained values for the responses 25.694, and 6.393 for Retention Time and Asymmetry respectively specify an adequate signal and these models can be used to navigate the design space.

Table 4: Box-Behnken design with responses

Trail No	Run Order	% Organic Composition	Flow Rate	Organic Modifier	Retention Time	Theoretical Plates	Asymmetry
5	1	30	0.8	ACN	2.289	8647	1.2
7	2	30	1	ACN	2.291	8562	1.2
10	3	40	0.8	Methanol	2.457	7988	1.2
17	4	40	0.9	ACN	2.290	8754	1.0
4	5	50	0.9	Methanol	2.319	8769	1.1
16	6	40	0.9	ACN	2.290	8754	1.0
12	7	40	1	Methanol	2.318	8669	1.0
13	8	40	0.9	ACN	2.290	8754	1.1
15	9	40	0.9	ACN	2.290	8754	1.1
11	10	40	1	Methanol	2.318	8669	1.0
3	11	30	0.9	Methanol	2.321	8869	1.2
14	12	40	0.9	ACN	2.290	8754	1.0
8	13	50	1	Methanol	2.321	8869	1.2
9	14	40	0.8	Methanol	2.310	8569	1.2
1	15	30	0.9	Methanol	2.321	8869	1.2
6	16	50	0.8	ACN	2.325	8216	1.1
2	17	50	0.9	Methanol	2.319	8769	1.1

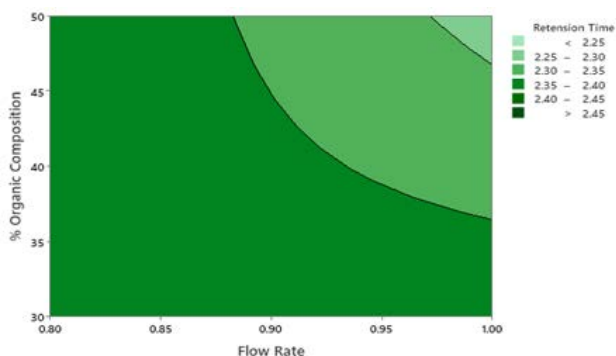


Figure 3: 2D Contour plots of retention time as a function of % organic composition in mobile phase and flow rate

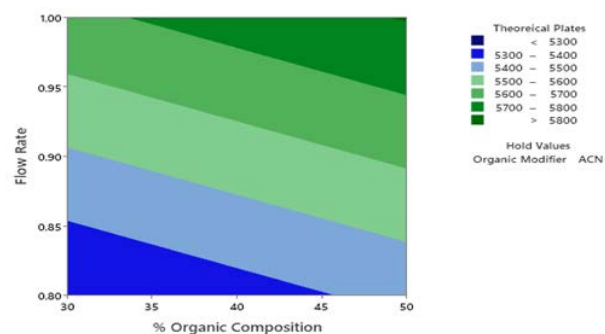


Figure 4: 2D Contour plots of theoretical plates as a function of % organic composition in mobile phase and flow rate

Table 5: ANOVA table for Retention Time using Box-Behnken design

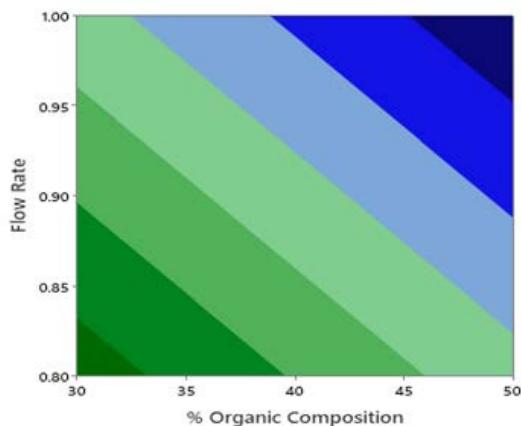
ANOVA for Response Surface Linear model						
Analysis of variance table [Partial sum of squares - Type III]						
Source	Sum of squares	degree of freedom	Mean square	F value	p-value	Inference
Model	3.13	5	1.01	67.13	<0.0001	Significant
A-OP	2.80	1	2.80	183.91	<0.0001	Significant
B-FR	0.0836	1	0.0837	5.51	0.031	Significant
C-OM	0.2403	1	0.2403	15.89	0.001	Significant
Residual	0.1961	11	0.0153			

Table 6: ANOVA table for Theoretical Plate using Box-Behnken design

ANOVA for Response Surface Linear model						
Analysis of variance table [Partial sum of squares - Type III]						
Source	Sum of squares	degree of freedom	Mean square	F value	p-value	Inference
Model	2.456E+05	3	8.186E+06	2.59	0.095	Not Significant
A-OP	1.941E+07	1	1.941E+07	6.17	0.023	Significant
B-FR	2.785E+06	1	2.785E+06	0.8870	0.361	
C-OM	2.357E+05	1	2.353E+06	0.7501	0.401	
Residual	4.083E+07	13	3.140E+06			

Table 7: ANOVA table for Asymmetry using Box-Behnken design

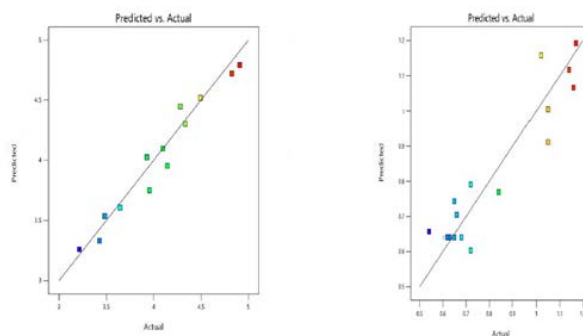
ANOVA for Response Surface Linear model						
Analysis of variance table [Partial sum of squares - Type III]						
Source	Sum of squares	degree of freedom	Mean square	F value	p-value	Inference
Model	0.7031	9	0.0783	5.31	0.019	Significant
A-OP	0.0701	1	0.0701	4.78	0.061	
B-FR	0.0251	1	0.0251	1.75	0.223	
C-OM	0.0363	1	0.0363	2.51	0.153	
AB	0.0890	1	0.0890	6.21	0.040	
AC	0.0759	1	0.0759	5.29	0.054	
BC	0.0651	1	0.0651	4.50	0.070	
A2	0.0531	1	0.0531	3.71	0.095	
B2	0.2681	1	0.2681	0.0657	0.803	Significant
C2	0.0007	1	0.0007	18.90	0.001	
Residual	0.1008	7	0.0134			

**Figure 5:** 2D Contour plots of asymmetry as a function of % organic composition in mobile phase and flow rate

Design validation

From the predicted versus actual plots of retention time

and asymmetry given in Figure 6, it was detected that the selected models for the respective responses were fit for the selected design as this plot specifies the uniform distribution of the data points around the 45° line. It was further proved from the ANOVA Tables 4 and 6 that the selected models were significant with $P < 0.05$ and apt for the design employed in this work

**Figure 6:** Predicted versus actual plots of Retention Time and Asym-

metry

Optimization by desirability functions approach

The optimized chromatographic conditions designated based on the desirability functions approach were mobile phase comprising of Acetonitrile: 0.02 N OPA (47:53 v/v) pumped at a flow rate 0.96 ml/min of gave the highest desirability of 0.900. The overlay contour plot shown in Figure 7 denotes the optimized combination of the three selected independent factors which gives the maximum desirability. To check this optimum set of conditions, three replicate injections of 100 µg/ml Ezetimibe were analyzed to determine if their observed responses were within the predicted range as shown in Table 8 and the corresponding optimized chromatogram was shown in Figure 8

Table 8: Responses of the optimized method

S. No.	Response variables	Predicted value	Actual value	Desirable range
1	Retention time (min)	2.879	2.795	2.49436–3.96364
2	Theoretical plates	7921.66	8345	5203.18–6423
3	Asymmetry	1.01	1.04	0.9-1.2

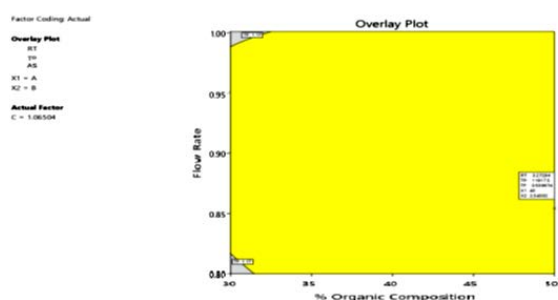


Figure 7: Overlay contour plot supported by responses

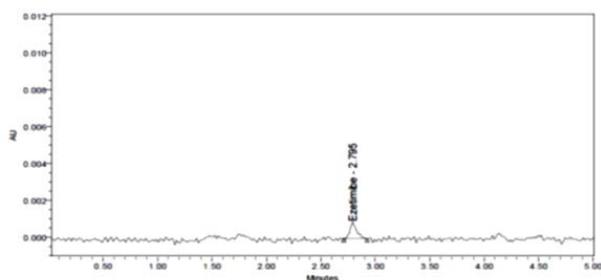


Figure 8: Chromatogram of the optimized method

Optimized chromatographic conditions suggested by DOE

Column: Phenomenex C18 (250 mm × 4.6 mm, 5 µm)

Mobile phase: 0.02 N OPA: Acetonitrile (53:47%)

Flow rate: 0.96 ml/min

Wavelength: PDA-UV detection at 232.6 nm

Column temperature: Ambient

Run time: 5 minutes

Method validation

The developed method was linear over the concentration range of 1.25 µg/ml-7.5 µg/ml with a correlation coefficient of 0.999. For the accuracy studies at 50%, 100% and 150% levels the recovery percentage of the drug was found to be within 98.42%-101.11%. System precision, Intermediate precision, and repeatability were carried out and the percent of RSD values were found to be less than 2%. LOD and LOQ values were found to be 0.02 µg/ml and 0.06 µg/ml. The robustness of the developed method was checked by making minor changes in the experimental conditions like flow rate, Percentage of organic composition, and Percent of RSD values for the peak area were found to be less than 2%. From the system suitability tests, the number of theoretical plates was found to be more than 2000 and the tailing factor was found to be less than 2. The summary of the method validation parameters was shown in Table 9. Forced degradation studies of Ezetimibe was performed in various conditions like acidic, basic, peroxide, thermal, photolytic, hydrolytic and reductive. Data of forced degradation studies were presented in Table 10.

Table 9: Results of the validation parameters

S No.	Parameters	Results
1	Linearity	
	Linearity range (µg/ml)	1.25-7.5
	Correlation coefficient	0.999
	Regression equation	y=43904.x+1683.5
2	Accuracy (% recovery)	
	50%, 100%, 150% levels	Between 98.42 and 101.11
3	Precision (% RSD of peak area)	
	System precision	1.2
	Repeatability	1.2
4	Sensitivity	
	LOD (µg/ml)	0.02
	LOQ (µg/ml)	0.06
5	Robustness (% RSD of peak area)	
	Flow rate (± 0.1 ml/min)	1.2
	Organic phase (± 10%)	0.45
	Temperature (± 5°C)	1.35
6	System suitability	
	Retention time (min)	2.795
	Tailing factor	1.21
	Plate count	8345

Table 10: Results of forced degradation studies

Drug	Degradation Condition	% Recover	% Drug Degraded
Ezetimibe	Acid	94.27	5.73
	Alkali	95.12	4.88
	Oxidation	94.36	5.64
	Thermal	96.86	3.14
	UV	98.17	1.83
	Water	99.16	0.84

Conclusion

A simple, precise, robust, and accurate RP-HPLC method

was created for the estimation of Ezetimibe by using the Design of Experiments approach. Box-Behnken Design with three factors of three levels was chosen as the optimization design for the present study. The critical method parameters selected for optimization were percentage of organic content in the mobile phase, organic modifier, and flow rate. The critical quality attributes are retention time, theoretical plates, and Asymmetry. Optimized chromatographic conditions advised by the desirability functions approach comprised of mobile phase 0.02 N OPA (53%): Acetonitrile (47%), which is pumped at a flow rate of 0.96 ml/min gave the highest desirability of 0.9. The retention time of the drug was found to be 2.795 min. Theoretical plates and Asymmetry was within the limits. The validation of the developed method was done as per ICH Q2 (R1) guidelines. The application of RSM offers a better perception of method development and robustness testing. In various stress conditions, degradation studies were also performed.

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References

1. M. V. Heek, H. Davis, Pharmacology of Ezetimibe, *Eurn Hrt J Splmnts*, 4 (2002), J5–J8.
2. B. Shailendra Patel, Ezetimibe: A novel cholesterol-lowering agent that highlights novel physiologic pathways, *Currn Card Rprts*, 6(2004), 439-442.
3. K. S. Prabhat, K. B. Pawan, K. S. Sushant, J. Deepti, Validated RP- HPLC method for estimation of Ezetimibe in different tablet dosage form, *Int J Pharm Sci*, 1(2009), 174-181.
4. P. Kumar, Y. Ahmad, G. Amitav, A stability indicating RP-HPLC method development for determination of Ezetimibe in tablet dosage form, *Der Pharma Chemica*, 4(2012), 1296-1304.
5. P. Ghanshyam, S. Ragvendra, S. Gautham, D. Dinesh, Development and validation of a new reversed-phase HPLC method for the determination of Ezetimibe in pharmaceutical dosage forms, *Indn J Pharm Edu and Res*, 47(2013), 7-12.
6. T. A. Wani, A. Ahmad, S. Zargar, N. Y. Khalil, I. A. Darwish, Use of response surface methodology for development of new micro well-based spectrophotometric method for determination of atrovastatin calcium in tablets, *Chem Cen J*, 6 (2012), 134.
7. M. Deepa, K. Ravindra Reddy, S. V. Satyanarayana, A review on quality by design approach for analytical method development, *J Pharm Res*, 11 (2017), 272-77.
8. ICH Harmonised Tripartite Guideline: Validation of Analytical Procedures: Text and Methodology Q2 (R1), Current STEP 4 Version. Geneva: International Conference on Harmonisation; 2005.
9. M. Blessy, D. R. Patel, N. P. Prajapati, Y. K. Agarwal, Development of forced degradation and stability indicating studies of drugs-a review, *J Pharm Anlys*, 4 (2014), 159-65.
10. M. Deepa, K. Ravindra Reddy, S. V. Satyanarayana, A review on quality by design approach for analytical method development, *J Pharm Res*, 11(2017), 272-77.
11. G. E. P. Box, D. W. Behnken, Some new 3 level designs for the study of quantitative variables, *J Technometrics*, 2(1960), 455-75.